ELSEVIER

Contents lists available at ScienceDirect

Environmental Pollution

journal homepage: www.elsevier.com/locate/envpol



Analysis of a Farquhar-von Caemmerer-Berry leaf-level photosynthetic rate model for *Populus tremuloides* in the context of modeling and measurement limitations

Kathryn E. Lenz ^{a,*}, George E. Host ^b, Kyle Roskoski ^b, Asko Noormets ^{c,1}, Anu Sôber ^d, David F. Karnosky ^c

A photosynthetic rate model is parameterized for Populus tremuloides and evaluated based on its ability to predict dependent as well as independent data.

ARTICLE INFO

Article history: Received 11 August 2009 Accepted 17 August 2009

Keywords:
Photosynthetic rate model
Populus tremuloides
Parameterization
Validation

ABSTRACT

The balance of mechanistic detail with mathematical simplicity contributes to the broad use of the Farquhar, von Caemmerer and Berry (FvCB) photosynthetic rate model. Here the FvCB model was coupled with a stomatal conductance model to form an $[A,g_s]$ model, and parameterized for mature *Populus tremuloides* leaves under varying CO_2 and temperature levels. Data were selected to be within typical forest light, CO_2 and temperature ranges, reducing artifacts associated with data collected at extreme values. The error between model-predicted photosynthetic rate (A) and A data was measured in three ways and found to be up to three times greater for each of two independent data sets than for a base-line evaluation using parameterization data. The evaluation methods used here apply to comparisons of model validation results among data sets varying in number and distribution of data, as well as to performance comparisons of $[A,g_s]$ models differing in internal-process components.

© 2009 Elsevier Ltd. All rights reserved.

1. Introduction

The widely accepted, steady-state photosynthetic rate model of Farquhar, von Caemmerer and Berry (1980), the FvCB model, relates C_3 leaf gas exchange data to underlying limitations to photosynthesis at the leaf-tissue level due to the activity of Rubisco, regeneration of RuBP and stomatal conductance. The model achieves a useful balance between mechanistic detail and mathematical simplicity. Various modifications to the model have been developed in order to extend its responsiveness to a wide range of environmental conditions (Hikosaka et al., 2006; Rogers and Humphries, 2000) water and nutrient stress (Dewar, 2002; Kubiske et al., 2002), and elevated CO_2 and O_3 concentrations (Karnosky et al., 2003; Kull et al., 1996; Martin et al., 2000, 2001; Reich, 1983).

As noted by Farquhar, von Caemmerer and Berry (2001), the FvCB model does not include all mechanisms contributing to

photosynthetic rate. Instead it represents an intelligent, simplified blend of processes occurring at the chloroplast level. Extrapolation of the model to the leaf-level, however, can result in erroneous assumptions about homogeneity of photosynthetic activity throughout the leaf, gas conductances and other internal processes (Flexas et al., 2008; Schurr et al., 2006). In order to predict photosynthetic rate from external rather than internal CO₂ concentration, the FvCB model is coupled with a stomatal conductance model, often of the Ball-Berry (Ball et al., 1987) or Leuning (Leuning, 1995) type, forming an [A,g_s] model.

Different strategies for calibrating the non-linear FvCB model, with its interdependent parameters, can lead to different sets of parameter values (see e.g. Sharkey et al., 2007). For example, Dubois et al. (2007) showed that, using standard parameterization methods, the assumed value of the A/C_i transition point alone can significantly influence the resulting estimates of $V_{\rm cmax}$, $J_{\rm max}$ and $R_{\rm d}$. (See the Appendix for symbol definitions and units.) Moreover, parameters for these components are typically determined as optimal regression fits to data and such fits can be sensitive to data distribution, inaccuracies and bias.

Unrecognized limitations and bias of instrumentation can also be misleading (Long and Bernacchi, 2003; Long et al., 1996). Estimates of respiration, $\rm CO_2$ compensation point, $V_{\rm cmax}$ and $J_{\rm max}$ can all vary significantly with instrumentation and techniques

^a Department of Mathematics and Statistics, University of Minnesota Duluth, 10 University Drive, Duluth, MN 55812, USA

b Natural Resources Research Institute, University of Minnesota Duluth, 5013 Miller Trunk Highway, Duluth, MN 55811, USA

^c School of Forestry Resources and Environmental Science, Michigan Technological University, 1400 Townsend Drive, Houghton, MI 49931, USA

^d Institute of Ecology and Earth Sciences, University of Tartu, 40 Lai Str. 51005, Tartu, Estonia

^{*} Corresponding author. Tel.: +1 218 726 7216; fax: +1 218 726 8399. E-mail addresses: klenz@d.umn.edu (K.E. Lenz), ghost@nrri.umn.edu (G.E. Host), kroskosk@d.umn.edu (K. Roskoski), anoorme@ncsu.edu (A. Noormets), anu.sober@ut.ee (A. Sôber).

¹ Current address: Department of Forestry and Environmental Resources, North Carolina State University and Southern Global Change Program, USDA Forest Service, Raleigh, NC 27695-7260, USA.

(Bernacchi et al., 2002; Ethier and Livingston, 2004; Singsaas et al., 2003). Data collected at extreme C_a levels within a gas exchange chamber are proportionately more affected by measurement error, imprecision and CO_2 leakage and diffusion than when chamber C_a levels are close to external C_a levels (Long and Bernacchi, 2003; McDermitt et al., 1989; Rodeghiero et al., 2007). Because of the technique by which A is measured, CO_2 leaking or diffusing into the chamber at low chamber C_a reduces apparent A, while CO_2 leaking out of the chamber at high chamber C_a increases apparent A. Also the use of C_i in place of C_c introduces bias in estimation of V_{cmax} and J_{max} (Singsaas et al., 2003), though the difference between C_i and C_c is difficult to predict (Flexas et al., 2007, 2008).

Measurements to estimate J_{max} are often made at chamber $C_{\text{a}} = 2000$ ppm to ensure that CO_2 is saturating and hence photosynthesis is RuBP regeneration limited. However at very high cuvette C_{a} there can be measurement problems related to stomatal closure in illuminated leaves (Niinemets et al., 1999). Also, phosphate availability may curb RuBP regeneration (Niinemets et al., 1999; Sharkey, 1985), leading to underestimation of J_{max} .

This study parameterized and validated a coupled $[A,g_s]$ model using leaf-level A/C_i data with the goal of predicting A at growth C_a levels while seeking to minimize measurement effects and unmodeled photosynthetic rate dynamics. Validation was based on the $[A,g_s]$ model's predictions of measured A values for dependent (i.e. parameterization) and independent data sets. Parameterization and validation were limited to leaf-level photosynthetic rate for healthy, mature P. tremuloides Michx. sun leaves with environmental conditions typical of aspen forests, in the absence of water and nutrient stress.

2. Methods of model development

2.1. Mathematical model

In Farquhar et al. (1980) steady-state C_3 leaf photosynthetic carbon assimilation rate A is driven by intercepted light, CO_2 , temperature, and humidity as well as internal leaf processes. Photosynthetic rate is assumed to be limited either by Rubisco-catalyzed carboxylation, the regeneration of RuBP controlled by electron transport rate, or the regeneration of RuBP controlled by the rate of triose-phosphate utilization, TPU. No evidence of TPU limitation was found in the data (Long and Bernacchi, 2003; von Caemmerer, 2000). Thus $A = \min \{A_c, A_j\}$, where Rubisco-limited A is given by

$$A_{\rm C} = \frac{V_{\rm c,max}(C_{\rm i} - I^*)}{C_{\rm i} + K_{\rm m}} - R_{\rm d},\tag{1}$$

where $K_{\rm m}=K_{\rm c}(1+\frac{O_{\rm i}}{K_{\rm o}})$, and RuBP regeneration limited A is given by

$$A_{j} = \frac{J(C_{i} - \Gamma^{*})}{4.5C_{i} + 10.5\Gamma^{*}} - R_{d}$$
(2)

As in Harley and Tenhunen (1991), dark respiration was expressed as an Arrhenius function of temperature of the form

$$R_{\rm d} = \exp(c)\exp(-\Delta H_{\rm a}/(RT_{\rm k})) \tag{3}$$

The temperature dependencies for Γ^* and K_m were drawn from Bernacchi et al. (2002). The parameters $V_{\rm cmax}$ and $J_{\rm max}$ were modeled as in Harley and Tenhunen (1991), Harley et al. (1992b) and Medlyn et al. (2002) by peaked temperature functions of the form

$$P(T_{k}) = P_{\text{opt}} \left(\frac{\Delta H_{\text{d}} \exp\left(\frac{\Delta H_{\text{a}}(T_{k} - T_{\text{opt}})}{T_{k}RT_{\text{opt}}}\right)}{\Delta H_{\text{d}} - \Delta H_{\text{a}} \left(1 - \exp\left(\frac{\Delta H_{\text{d}}(T_{k} - T_{\text{opt}})}{T_{k}RT_{\text{opt}}}\right)\right)} \right), \tag{4}$$

where $P(T_k)$ is the value of the parameter at temperature T_k , P_{opt} is the maximum value of the parameter and T_{opt} is the temperature at which P_{opt} is achieved.

The rate of electron transport, J, was related to J_{max} according to

$$\Theta_{PSII}J^2 - (Q_2 + J_{max})J + Q_2J_{max} = 0$$
 (5)

The temperature dependences of Θ_{PSII} and Q_2 were those of Bernacchi et al. (2003) for growth temperature 14 °C.

As implemented in LI-6400 (equations from Farquhar and Sharkey, 1982; and Farquhar and von Caemmerer, 1982),

$$C_i = \frac{(g_{tc} - E/2000)C_s - A}{g_{tc} + E/2000},\tag{6}$$

for $C_s=\frac{F}{100S}C_r-A\bigg/\frac{F}{100S}+0.001E$, where F is air flow rate and S is leaf surface area inside the cuvette. In this study, $g_{tc}=\left(\frac{1.6}{g_s}+\frac{1.37(0.5)}{1.42}\right)^{-1}$, where g_s has the form

$$g_{s} = \frac{a_{1}(A + R_{d})}{C_{i}(1 + \frac{D_{i}}{D_{i}})} \tag{7}$$

This is a variation of the basic Ball-Berry-Leuning g_s model (Leuning, 1995).

2.2. Parameterization data

This study differs from previous work in the selection criteria developed and applied to data prior to parameterization. The intent was to use real-world data sets filtered to avoid or reduce uncertainty due to unmodeled photosynthesis dynamics associated with leaf-age, leaf-health, low stomatal conductance, photoinhibition, water stress or nutrient availability. The data were also filtered to reduce the effects of potential instrumentation-induced errors at very low or very high values of $C_{\rm r}$. Model components were parameterized primarily to high-light data with $C_{\rm r}$ close to $C_{\rm a}$ growth levels, where steady-state photosynthesis measurements are most likely representative of photosynthesis in the field.

The data used for parameterization in this study were collected as part of the Aspen FACE project (Dickson et al., 2000; Karnosky et al., 2003), located at 45°30' N, 89°30′ W. Leaf-level data were collected from P. tremuloides clone 216 growing in open-air conditions with exposure to elevated (550 ppm) and ambient (360 ppm) levels of CO2 during the 1998 and 2000 growing seasons. All measurements were made using an LI-6400 portable open gas exchange photosynthesis system (LI-COR, 1998, 2006). Only data for leaves within a Leaf Plastochron Index (LPI) range of approximately 9-26 were used. This range includes predominately recently mature and mature leaves. Kull et al. (1996) placed leaf maturation, based on chlorophyll content, at about LPI 8, while Noormets et al. (2001a) placed leaf maturation at about LPI 11. Because photosynthetic rate capacity may diminish in over-mature leaves and in leaves damaged by ozone (Kim and Lieth, 2003; Reich, 1983; Schultz, 2003; Niinemets et al., 2005), data for clearly over-mature or damaged leaves were excluded according to LPI and cut-off values for A. For $Q \ge 1000 \ \mu mol \ m^{-2} \ s^{-1}$, A was required to be at least 15 μ mol m⁻² s⁻¹ when $C_r > 300$ ppm and at least 20 when $C_{r} > 400$ ppm. Data for which measured g_{s} averaged less than 0.3 when C_{i} was between 145 and 500 ppm were also excluded as low g_s measurements could occur for various reasons associated with unmodeled leaf-tissue behavior (Farquhar and Sharkey, 1982; Leuning, 1995; Mott and Buckley, 2000; Noormets et al., 2001b).

The full parameterization data set, the [A,gs]-data, was the union of the first three limited data sets listed in Table 1. A large proportion of the [A,gs]-data were for C_r levels between 340 and 560 ppm, since predicting A for ambient growth conditions was of highest priority. All of the data for $C_r < 400$ ppm was for leaves grown at daytime C_a close to 350 ppm and all of the data for $C_r > 400$ ppm was for leaves grown under elevated day-time C_a , about 550 ppm. The A_c -data consisted of the FACE-1998A data for which $C_r \le 360$ ppm and the FACE-1998B data for which $C_r \le 360$ ppm and $C_r \le 360$ ppm

The $V_{\rm cmax}$ parameterization data set (V-data) consisted of the $A_{\rm C}$ -data for which $C_{\rm r}$ was approximately 250–530 ppm. This included FACE-1998_A data for $C_{\rm r}$ between 340 and 360 ppm and FACE-1998_B $A/C_{\rm i}$ data for which $C_{\rm i}$ was nearly as high as the apparent transition point or the next $C_{\rm i}$ measurement below that. These values of $C_{\rm i}$ increased with temperature.

Table 1Parameterization and validation data sets, with external variable ranges.

Name	R _d measured	C _a ppm	Q μmol m ⁻² s ⁻¹	Temp °C	RH %	Reference
Parameterizati	Parameterization data sets					
FACE-1998 _A	no	360, 550	1000-1500	24-36	42-75	Noormets et al., 2001a
FACE-2000	no	1500	0-500	21-29	58-77	Noormets et al., 2010
FACE-1998 _B Validation data	yes a sets	50-410	1000-1500	19–36	23-81	
BOREAS TE-12	yes	70–400	0-2100	15–37	15-33	Arkebauer, 1998
FACE-1999	no	360, 560	1188-1300	17-35	46-77	Noormets et al., 2001b

For parameterization of J_{max} , rather than using A/C_i data in the flat region of the curve well above the transition point, we used data at or near the transition point, where the slope was greater. The J-data consisted of the high-light ($Q \ge 1000 \ \mu \text{mol m}^{-2} \ \text{s}^{-1}$) portion of the A_J -data set. Although it is possible that some J-data points should have been in the A_C -data instead, there was the benefit of targeting parameterization of A_J directly to a realistic environmental CO_2 concentration range.

2.3. Parameterization procedure

The full parameterization of the $[A,g_s]$ -model was achieved in stages by fitting model components to data using non-linear regression (NLIN Procedure, SAS 8.1; The SAS Institute, Inc., Cary, NC, USA). Prior to parameterization the data were corrected for measurement error due to respiring tissue beneath the gasket as described by Pons and Welschen (2002) and validated for largely heterobaric leaves (Jahnke, 2001). This gasket error occurs when measured respiration is divided by the leaf area within the cuvette chamber, ignoring the portion (assumed to be half) of the leaf-tissue under the gasket which also respired into the chamber. Underestimation of respiring leaf area results in over-estimation of R_d .Based on the LI-6400 leaf cuvette area and gasket thickness, the assumed gasket error was corrected by dividing measured R_d by 1.59. The corresponding corrections to the A and C_i data were made by adding the R_d correction to A and then recalculating C_i using Eq. (6).

The Arrhenius function (3) was fit to the corrected R_d data. Next, V_{cmax} data points were generated using the equation

$$V_{\text{cmax}} = \frac{(C_i + K_m)(A + R_d)}{(C_i - \Gamma K_m)},$$
(8)

the V-data, and the $R_{\rm d}$, T^* and $K_{\rm m}$ temperature functions. Rather than compute $V_{\rm cmax}$ as the slope of a regression line through three to five points on the $A/C_{\rm i}$ curve, $V_{\rm cmax}$ was calculated directly from (8). This allowed inclusion of the FACE-1998_A data, where measurements were taken at only a single $C_{\rm r}$ value per leaf.

The temperature function (4) was fit to the $V_{\rm cmax}$ points. The value of $\Delta H_{\rm d}$ was fixed and then $T_{\rm opt}$, $V_{\rm cmax,opt}$ and $\Delta H_{\rm d}$ were derived using SAS. As $\Delta H_{\rm d}$ was increased incrementally from 150 000 to 700 000 J mol $^{-1}$ the shape of the resulting peaked function changed from a gently sloping, symmetric bell to an asymmetric one resembling an Arrhenius curve below $T_{\rm opt}$. Primarily, increasing $\Delta H_{\rm d}$ increased the value of $V_{\rm cmax,opt}$ and the steepness of descent above $T_{\rm opt}$.

The *J* points were generated from the equation

$$J = \frac{(4.5C_i + 10.5\Gamma^*)(A + R_d)}{(C_i - \Gamma^*)},$$
(9)

the J-data and the $R_{\rm d}$ and \varGamma^* temperature functions. From the J points, $J_{\rm max}$ points were computed according to (5). The temperature dependency function (4) was then fit to the $J_{\rm max}$ points.

The g_s model parameter a_1 was determined by a linear regression fit of the g_s data to the quantity $\frac{A+R_d}{C_i(1+\frac{D_s}{2})}$ using the R_d temperature function and the A, C_i and D_s data in the $[A,g_s]$ -data set for which g_s was at least 0.3 mol m⁻² s⁻¹.

2.4. Model validation

Common statistical procedures programmed into spreadsheet software were used for model validation. The parameterized $R_{\rm d}$, $A_{\rm c}$, $A_{\rm j}$ and $[A,g_{\rm s}]$ models were first evaluated against the $[A,g_{\rm s}]$ -data. These data were considered dependent because modeling components were parameterized to subsets of the $[A,g_{\rm s}]$ -data set. These initial evaluations served as benchmarks for comparisons with independent data.

In addition to visual inspection of data values $X_1, X_2, ... X_n$ plotted against their corresponding model-predicted values $Y_1, Y_2, ... Y_n$, goodness of fit was assessed based on the slope and R^2 value of the optimal least squares regression line, the root mean squared error ε and the root mean squared relative error ε _r where

$$\varepsilon = \frac{1}{n} \sqrt{\sum_{i=1}^{n} (Y_i - X_i)^2},\tag{10}$$

$$\varepsilon_r = \frac{1}{n} \sqrt{\sum_{i=1}^{n} \left(\frac{Y_i - X_i}{X_i}\right)^2} \tag{11}$$

The optimal regression line is of the form $y=\alpha x$, where α minimizes the error $\frac{1}{n}\sqrt{\sum_{i=1}^{n}(Y_i-mX_i)^2}$ over all possible slopes m. The quantities ε and ε_r directly measure the error in a model's ability to predict the data points $X_1,X_2,...X_n$.

Two of the data sets in this study had A values ranging from very small to large, but the third data set had only upper-range values of A ($A \ge 15$ µmol m⁻² s⁻¹). For this reason and to avoid comparing relative error for small A with relative error for large A, we used ε , but not ε _r, to compare the [A,gs] model's error in predicting A among whole data sets. Both ε and ε _r were reasonable choices for

comparison of the [A,gs] model's error in predicting A over the limited range $A \ge 15~\mu mol~m^{-2}~s^{-1}$.

2.5. Validation data

Table 1 lists the independent model validation data sets. The BOREAS TE-12 data were collected from intact leaves of P. tremuloides during in 1994 and 1995 at the BOReal Ecosystem-Atmosphere Study near Prince Albert, Saskatchewan at 53.2° N (Arkebauer, 1998). An LI-6200 CO₂ infrared gas analyzer system was used in closed-circuit mode (LI-COR, 1990; McDermitt et al., 1989) with a 14 cm² chamber window. The FACE-1999 data were collected in 1999 at the FACTS-II site with an LI-6400 portable open gas exchange system and were measured only at growth C_a levels, approximately 350 and 550 ppm.

In order to validate the model for healthy, mature, sun leaves, the data were filtered in the same manner as the parameterization data. This resulted in data sets for leaves within an LPI range of 9–26, $Q \geq 1000~\mu \text{mol m}^{-2}~\text{s}^{-1}$, and $A \geq 15~\mu \text{mol m}^{-2}~\text{s}^{-1}$ when $C_r > 300~\text{ppm}$. Data for which g_s averaged less than 0.3 mol m $^{-2}~\text{s}^{-1}$ when C_l was between 145 and 500 ppm were excluded. While the BOREAS TE-12 data were distributed fairly evenly from very low to high values of A, the FACE-1999 data had only $A \geq 15~\mu \text{mol m}^{-2}~\text{s}^{-1}$.

Additional comparisons were made with Niinemets et al. (1999) LI-6262 open gas exchange system (LI-COR Inc., Lincoln, NE, USA) data collected for upper canopy *Populus tremula* L. leaves in Estonia (latitude 58°22'N). Comparison of $R_{\rm d}$ results were also made with *P. tremuloides* data from the Aspen FACE site in 2000 (Davey et al., 2004) and 2001 (Singsaas et al., 2003).

3. Results

3.1. R_d , V_{cmax} and J_{max} models

The $R_{\rm d}$ model is comparable to the Niinemets et al. (1999) $R_{\rm d}$ model and the BOREAS TE-12 $R_{\rm d}$ data (Fig. 1, Table 2). The Niinemets et al. (1999) $R_{\rm d}$ model was fit to data taken over a temperature range of 20–50 °C. Our model predicts $R_{\rm d}=1.88~\mu{\rm mol~m^{-2}~s^{-1}}$ at 25 °C. In comparison, LI-6400 $R_{\rm d}$ data at 25 °C reported by Singsaas et al. (2003), when corrected for the gasket error, range from 1.95 to 2.75 $\mu{\rm mol~m^{-2}~s^{-1}}$, with mean value 2.35 $\mu{\rm mol~m^{-2}~s^{-1}}$. The range of $R_{\rm d}$ for aspen published in Davey et al. (2004) is 1.31–1.47 $\mu{\rm mol~m^{-2}~s^{-1}}$ at 25 °C. The Davey et al. (2004) study instrumentation had a cuvette large enough to enclose an entire leaf, avoiding the gasket error.

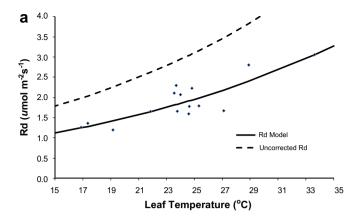
Because it is proportional to R_d , the R_d gasket error increases with temperature. Thus correcting the error reduces the apparent rise in R_d , and consequently Γ^* , with temperature. It was found that without the R_d correction, Γ^* increased with the slope of Γ^* in Bernacchi et al. (2001) but had lower magnitude. With the R_d correction, Γ^* increased with the slope of Γ^* in Bernacchi et al. (2002) and had similar magnitude.

The $V_{\rm cmax}$ and J points were computed as described in Subsection 2.3 using Eqs. (8) and (9). These data and the $R_{\rm d}$ data were confirmed by comparison with $V_{\rm cmax}$, $R_{\rm d}$ and $J_{\rm max}$ points derived using slope-intercept methods applied to (1) and (2) as described in von Caemmerer (2000).

The V_{cmax} temperature function was fit to the V_{cmax} data (Fig. 2, Table 2). Its value at 25 °C, about 95 μ mol m⁻² s⁻¹, is between the two Singsaas et al. (2003) estimates of V_{cmax} at 25 °C for leaves grown under an ambient CO₂ level of 350 ppm ($V_{cmax}=80~\mu$ mol m⁻² s⁻¹ estimated from A/C_i data, $V_{cmax}=102~\mu$ mol m⁻² s⁻¹ estimated from A/C_c data).

An Arrhenius model fit the $V_{\rm cmax}$ data well. All data were apparently for temperatures below $T_{\rm opt}$. For the peaked function, a deactivation energy of $\Delta H_{\rm d}=500~000~{\rm J~mol}^{-1}$ resulted in a fit comparable to an Arrhenius model fit (not shown). Fixing $\Delta H_{\rm d}$ and then applying SAS to simultaneously solve for $T_{\rm kopt}$, $V_{\rm cmax,opt}$ and $\Delta H_{\rm a}$ resulted in an estimation for $T_{\rm kopt}$ of 38 °C.

Often only A/C_i data well below the cross-over region are used for fitting A_c , as estimation of $V_{\rm cmax}$ could be compromised by accidental inclusion of RuBP regeneration-limited data. Indeed, Dubois et al. (2007) demonstrated that when A_c and A_i are fitted



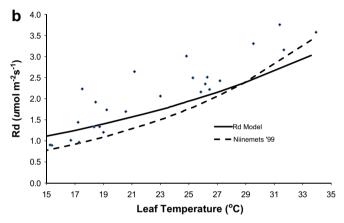


Fig. 1. a) Gasket-corrected R_d data, with R_d model fits to corrected and uncorrected data; b) the R_d model, the Niinemets et al. (1999) R_d model and the BOREAS TE-12 R_d data.

separately using points in or near the cross-over region, the estimates of $V_{\rm cmax}$ and $J_{\rm max}$ can be highly sensitive to the assumed value of the transition point.

In this study, $V_{\rm cmax}$ estimates computed from data near the transition point on the A/C_i curve, for C_i about 300–500 ppm, were nearly the same as the $V_{\rm cmax}$ points obtained from data lower on the A/C_i curve, but not lower than 150 ppm. We hypothesized that using data where $50 < C_i < 100$ ppm would result in lower $V_{\rm cmax}$ points due to CO_2 leakage into the sensor chamber, as well as unmodeled mesophyll conductance (Singsaas et al., 2003; Ethier and Livingston, 2004). However, instead we found only greater variability among the resulting $V_{\rm cmax}$ points.

The J_{max} points and temperature function were compared with J_{max} published in Niinemets et al. (1999) for P. tremula (Fig. 3, Table 2). Both J_{max} function values are about 170 μ mol m $^{-2}$ s $^{-1}$ at 25 °C, which is lower than the Singsaas et al. (2003) J_{max} values of 216 μ mol m $^{-2}$ s $^{-1}$ (for 550 ppm growth CO $_2$) and 230 μ mol m m $^{-2}$ s $^{-1}$ (for 350 ppm growth CO $_2$) estimated from A/C_i data, and 200 μ mol m $^{-2}$ s $^{-1}$ (for 550 ppm growth CO $_2$) and 217 μ mol m $^{-2}$ s $^{-1}$ (for 350 ppm growth CO $_2$), estimated from A/C_c data.

Table 2 Parameters for temperature models of $R_{\rm d}$, $V_{\rm cmax}$, and $J_{\rm max}$ assuming infinite $g_{\rm m}$.

	С	$P_{ m opt} \mu m mol \ m^{-2} s^{-1}$	$\Delta H_{\rm a}$ J mol $^{-1}$	$\Delta H_{ m d}$ J $ m mol^{-1}$	T _{opt} °C
R _d fit to FACE-1998 _B data	16.6	_	39 504	_	_
R _d Niinemets et al. (1999)	23.93	_	57 920	-	-
$V_{\rm cmax}$ fit to A/C_i V-data	-	233.53	59 808	500 000	38
J_{max} fit to $A/C_i J$ -data	-	250.62	33 228	500 000	36
J _{max} Niinemets et al. (1999)	-	380.00	70 600	285 800	34

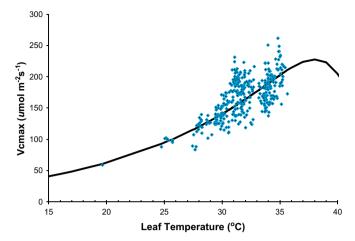


Fig. 2. The $V_{\rm cmax}$ model and the $V_{\rm cmax}$ data, assuming infinite $g_{\rm m}$.

3.2. Parameterized gs and [A,gs] models

The parameterized g_s model only coarsely predicted the g_s data. Nonetheless, it performed well as a feedback component within the [A,g_s] model. It was expected that the FvCB model would predict the A data better than the [A,g_s] model, since the FvCB model responds to C_i data while the [A,g_s] model responds to C_a . However, for the dependent [A,g_s]-data, predictions of A by the two models differed by no more than 2 μ mol m⁻² s⁻¹ when compared over the full range of data, and much less for $A < 15 \mu$ mol m⁻² s⁻¹.

Analysis of the $[A,g_s]$ model's ability to predict A for the $[A,g_s]$ -data (Fig. 4, Table 3) indicated base-line performance limitations due to some combination of data inaccuracies, parameterization techniques and unmodeled processes contributing to leaf-level photosynthetic rate. Validation with BOREAS TE-12 data (Fig. 5, Table 3) and FACE-1999 data (Fig. 6, Table 3) gave evidence of the $[A,g_s]$ model's ability to predict A for independent data.

Some of the error in predicting the BOREAS TE-12 data may be due to tree age, growth temperature and other environmental differences (Hikosaka et al., 2006). In contrast, the FACE-1999 data were collected at the same site as the parameterization data, only one year later and with water and nutrients not limiting. Thus one might expect better validation results for the FACE-1999 data than for the BOREAS TE-12 data.

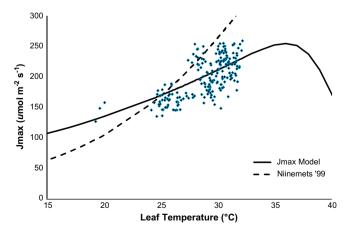


Fig. 3. The J_{max} model, the J_{max} data and the Niinemets et al. (1999) J_{max} model, assuming infinite g_{m} .

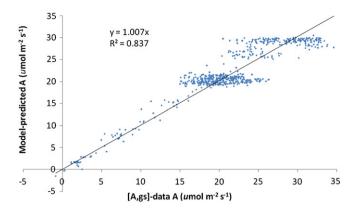


Fig. 4. A predicted by the [A,g_s] model versus LI-COR generated A in the [A,g_s]-data.

Both the regression line slope and R^2 values for the FACE-1999 validation were worse than for the BOREAS TE-12 validation (Table 3, column two). However, comparison of these measurements alone can be misleading because the A values in the BOREAS TE-12 data range from very low to high, while the FACE-1999 data have only $A \geq 15$ μ mol m⁻² s⁻¹. Limiting the data ranges to $A \geq 15$ μ mol m⁻² s⁻¹ for both the [A,g_s] and BOREAS TE-12 data sets resulted in R^2 values comparable to the R^2 value for the FACE-1999 data set (Table 3, third column).

In terms of ε_r for $A \ge 15 \ \mu mol \ m^{-2} \ s^{-1}$, the [A,g_s] model predicted the two independent data sets about equally well and with about 3 times the error than for the dependent data (Table 3 fifth column).

The optimal regression line slope suggests that the [A,g_s] model over-predicts the FACE-1999 A data by a factor of 1.219, but the low R^2 value reduces the certainty of this conclusion. The ε measure provides supporting evidence as follows. Let $X_1, X_2, \ldots X_n$ denote the FACE-1999 A data points and $Y_1, Y_2, \ldots Y_n$ denote the [A,g_s] model's predictions of these A values. The optimal regression line error (see Subsection 2.4) is

$$\varepsilon_{\text{opt}} = \frac{1}{n} \sqrt{\sum_{i=1}^{n} (Y_i - 1.219X_i)^2} = 0.131 \ \mu \text{mol m}^{-2} \text{s}^{-1}$$
(12)

This is about $2/5\epsilon$. Algebraic manipulation of (12) shows that if the model is scaled so that A=1/1.219 min $\{A_c,A_j\}$, it predicts the FACE-1999 A data with $\epsilon=0.1083$ µmol m⁻² s⁻¹. This is about as well as the $[A,g_s]$ model, without scaling, predicts the dependent $[A,g_s]$ -data.

3.3. The effects of g_m on apparent V_{cmax} and J_{max}

When V_{cmax} and J are estimated from A/C_i data as in Subsection 3.2, it is assumed that g_{m} is infinite, which is not realistic. To study

Table 3 Quantitative measures of the [A,g_s] model's ability to predict dependent and independent A data. The units for A and ε are μ mol m⁻² s⁻¹.

Data set	Regression all A data	Regression for $A \ge 15$	Root mean square error ε for all A	Root mean square relative error ε_r for $A \geq 15$
$[A,g_s]$	y = 1.007x $R^2 = 0.837$	y = 1.007x	0.105	0.00532
BOREAS TE-12	y = 1.000x	y = 0.953x $R^2 = 0.336$	0.136	0.01565
FACE-1999		$y = 1.219x$ $R^2 = 0.543$	0.310	0.01642

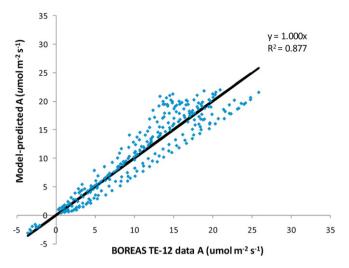


Fig. 5. Validation of the [A,g_s] model with the BOREAS TE-12 data.

the effects of $g_{\rm m}$ on $V_{\rm cmax}$ and $J_{\rm max}$ we assume that $C_{\rm c}=C_{\rm i}-A/g_{\rm m}$ as in Bernacchi et al. (2002), Flexas et al. (2007), Harley et al. (1992a) and Sharkey et al. (2007) and that $C_{\rm i}>\Gamma^*$. Replacing $C_{\rm i}$ in Eqs. (8) and (9) with $C_{\rm i}-A/g_{\rm m}$ expresses

 $V_{\rm cmax}$ and J at the site of Rubisco as functions of $g_{\rm m}$. Rearranging terms, we have

$$V_{cmax}(g_m) = \frac{(g_m - r_1)}{(g_m - r_2)} V_{cmax}(\infty),$$
 (13)

and

$$J(g_m) = \frac{(g_m - r_3)}{(g_m - r_2)} J(\infty), \tag{14}$$

where
$$r_1 = \frac{A}{C_i + K_m}$$
, $r_2 = \frac{A}{C_i - \Gamma^*}$ and $r_3 = \frac{A}{4.5C_i + 10.5\Gamma^*}$

with
$$0 < r_1 < r_2 < g_m$$
 and $0 < r_3 < r_2 < g_m$.

Since $V_{\rm cmax}(g_{\rm m})$ and $J_{\rm max}(g_{\rm m})$ have vertical asymptotes at $g_{\rm m}=r_2$, as the value of $g_{\rm m}$ is reduced toward r_2 , $V_{\rm cmax}(g_{\rm m})$ and $J(g_{\rm m})$ both increase without bound. Also their rates of change with respect to $g_{\rm m}$ become unbound.

 $V_{\rm cmax}(g_{\rm m})$ and $J(g_{\rm m})$ were investigated using the V-data and J-data for $g_{\rm m}=A/b$ for constant b ranging from 25 to 170 µmol mol⁻¹.

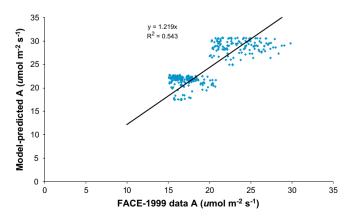


Fig. 6. Validation of the [A,gs] model with the FACE-1999 data.

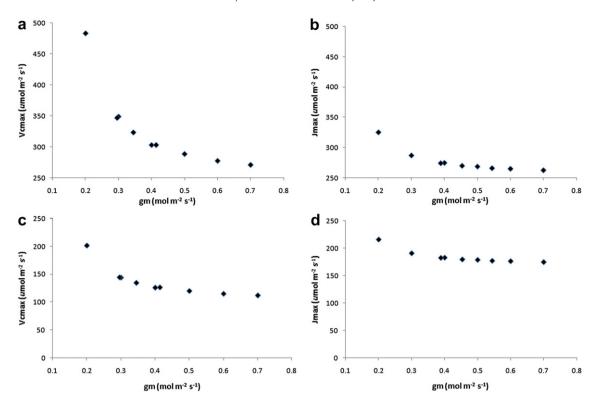


Fig. 7. For g_m assumed to be proportional to A in the V-data and J-data, \overline{g}_m plotted versus a) the parameter $V_{cmax,opt}$, b) the parameter $J_{max,opt}$, c) V_{cmax} at 25 °C, and d) J_{max} at 25 °C.

 $V_{\rm cmax}(g_{\rm m})$ and $J(g_{\rm m})$ points were generated from Eqs. (8) and (9) with $C_{\rm i}$ replaced by $C_{\rm c}=C_{\rm i}-A/g_{\rm m}=C_{\rm i}-b$. Temperature function fits of the $V_{\rm cmax}(g_{\rm m})$ and $J_{\rm max}(g_{\rm m})$ points were achieved by optimizing the coefficients $V_{\rm cmax,opt}$ and $J_{\rm max,opt}$ while maintaining the values of $\Delta H_{\rm a}$, $\Delta H_{\rm d}$ and $T_{\rm opt}$ given in Table 2. It was found that the difference between $V_{\rm cmax,opt}$ for $\overline{g}_{\rm m}=0.20$ mol m $^{-2}$ s $^{-1}$ (b=104 µmol mol $^{-1}$) and $V_{\rm cmax,opt}$ for $\overline{g}_{\rm m}=0.30$ mol m $^{-2}$ s $^{-1}$ (b=69 µmol mol $^{-1}$) was greater than the difference between $V_{\rm cmax,opt}$ for $\overline{g}_{\rm m}=0.30$ mol m $^{-2}$ s $^{-1}$ and $V_{\rm cmax,opt}$ for infinite $g_{\rm m}$ (Fig. 7). $V_{\rm cmax}(g_{\rm m})$ data values rapidly increased and dispersed as $\overline{g}_{\rm m}$ decreased below 0.20 mol m $^{-2}$ s $^{-1}$ and for $\overline{g}_{\rm m}<0.13$ mol m $^{-2}$ s $^{-1}$, within the range of $v_{\rm cmax}(g_{\rm m})$ values were huge and negative, some huge and positive and others were undefined.

4. Discussion

4.1. Empirical aspects of the V_{cmax} and J_{max} model components

Comparison of model-predicted A versus the $[A,g_s]$ -data A values verified that the limited V-data and J-data sets were sufficient for parameterizing $V_{\rm cmax}$ and $J_{\rm max}$ to the extent that $V_{\rm cmax}$ and $J_{\rm max}$ were satisfactory internal mathematical components in the $[A,g_s]$ model for predicting leaf-level A. Their effectiveness as components could be further explored by comparing the results in Table 3 with corresponding results obtained for other $[A,g_s]$ models with alternative $V_{\rm cmax}$ and $J_{\rm max}$ functions.

However, $V_{\rm cmax}$ and $J_{\rm max}$ were derived assuming infinite $g_{\rm m}$ and so do not accurately represent Rubisco dynamics (Niinemets et al., 1999; Singsaas et al., 2003). As shown in Subsection 3.3, assuming finite $g_{\rm m}$ results in higher estimates of both $V_{\rm cmax}$ and $J_{\rm max}$. However, even when finite $g_{\rm m}$ is estimated and included in the FvCB model using the relationship $C_{\rm c} = C_{\rm i} - A/g_{\rm m}$, $V_{\rm cmax}$ and $J_{\rm max}$ derived from $A/C_{\rm i}$ data can be significantly uncertain. We found that for high $g_{\rm m}$, where uncertainty in estimated $g_{\rm m}$ can be large (Harley et al., 1992a),

 $V_{\rm cmax}$ and $J_{\rm max}$ changed very little, as also reported by Flexas et al. (2007). However, when $g_{\rm m}$ is low $V_{\rm cmax}$ and J are very sensitive to uncertainty in $g_{\rm m}$. Accurate estimation of $g_{\rm m}$ may be difficult since, for a variety of C_3 species, $g_{\rm m}$ changes rapidly and significantly with CO_2 level during A/C_1 data collection (Flexas et al., 2007).

The values of $V_{\rm cmax}$ and $J_{\rm max}$ also depend on the values of Γ^* and $K_{\rm m}$, generally assumed to be species-independent for C_3 plants (von Caemmerer, 2000). Medlyn et al. (2002) observed sensitivity of $V_{\rm cmax}$ and $J_{\rm max}$ to Γ^* and $K_{\rm m}$. In this study, using Bernacchi et al. (2001) kinetics parameters, $V_{\rm cmax}$ assuming infinite $g_{\rm m}$ was larger than $V_{\rm cmax}$ for finite $\overline{g}_{\rm m}>0.7$ mol m⁻² s⁻¹ for the Bernacchi et al. (2002) kinetics parameters (Fig. 8). $V_{\rm cmax}$ increases because the

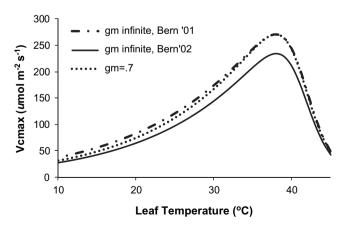


Fig. 8. The models for $V_{\rm cmax}$ assuming infinite $g_{\rm m}$ and $\overline{g}_{\rm m}=0.7$ mol m⁻² s⁻¹, both derived using the kinetics parameters from Bernacchi et al. (2002), compared to the model for $V_{\rm cmax}$ assuming infinite $g_{\rm m}$ derived using kinetics parameters from Bernacchi et al. (2001).

Bernacchi et al. (2001) Γ^* and $K_{\rm m}$ are larger at every temperature than the Bernacchi et al. (2002) I^* and $K_{\rm m}$. More generally, it is evident from Eq. (8) that with all other terms fixed, $V_{\rm cmax}$ increases with increasing $K_{\rm m}$. As I^* increases, the positive denominator $(C_i - \Gamma^*)$ in Eq. (8) decreases, which also increases $V_{\rm cmax}$. When C_i is replaced with C_c in Eq. (8), the same result is evident for $V_{\rm cmax}$ at the site of Rubisco. Thus erroneous conclusions may be drawn from comparison of $V_{\rm cmax}$ from studies using different I^* and $K_{\rm m}$ functions, even if all other aspects of the studies are the same.

4.2. Broader applicability of the analysis methods

The base-line and validation analysis methods of this study can be replicated using simple spreadsheet functions. They can be applied to evaluate and compare various parameterizations and alternative formulations of the $[A,g_s]$ model. For example, one could compare the performance of an [A,gs] model derived using simultaneous measurements of leaf gas exchange and modulated chlorophyll fluorescence (Long and Bernacchi, 2003; Sharkey et al., 2007) with an [A,g_s] model parameterized using traditional A/C_i techniques. Though the models may differ in source data and parameterization methods, they can be compared as long as they predict A from the same environmental inputs. Similarly, results of different mathematical formulations of components within the [A,g_s] model can be compared. The g_s model of Dewar (2002), for example, responds to leaf water potential as well as humidity. One could use these analysis methods to explore the extent to which substituting a Dewar (2002) g_s model for g_s in Eq. (7) improves an $[A,g_s]$ model's prediction of A data under various water-availability scenarios.

5. Conclusions

This study included parameterization, validation and analysis of a coupled $[A,g_s]$ model of photosynthetic rate for the most productive leaves of P. tremuloides. Restricting model parameterization data to limited ranges of environmental variables reduced uncertainty due to measurement limitations at very high and very low chamber C_a and unmodeled changes in internal conditions and mechanisms limiting photosynthetic rate. The model analysis incorporated elementary statistical computations programmed into spreadsheet software.

The analysis methods in this study apply more broadly, irrespective of parameterization techniques and the specific equations of [A,g_s] models. The base-line evaluation with dependent data quantifies the extent to which an [A,g_s] model, its mathematical structure in combination with its parameter values, is able to reproduce a larger body of data from which it was parameterized. This gives a context for assessment of independent validation results. The ε and ε_r measurements complement the graphical and optimal regression line information to facilitate comparison of model validation results for data sets varying in number and distribution of data values.

Acknowledgements

We gratefully acknowledge support from the USDA Northern Global Change Program Forest Service, Award #05-CA-11242343-036, the USDA Forest Service North Central Research Station Integrated Program, Agreement 03-JV-11231300-086, and the Office of Science (BER), U.S. DOE Grant No. DE-FG02-95ER62125. Additional support was provided by the University of Minnesota Duluth's Natural Resources Research Institute and Department of Mathematics and Statistics, and Michigan Technological University.

Appendix

Symbols

Α	Steady-state net rate of CO ₂ uptake per unit leaf area (µmol m ⁻² s ⁻¹)
A_{c}	A limited by Rubisco-catalyzed carboxylation (μmol m ⁻² s ⁻¹)
$A_{\rm j}$	A limited by the regeneration of RuBP controlled by electron
-	transport rate (μmol m ⁻² s ⁻¹)
a_1	Empirical coefficient (9.5)
С	Scaling constant (dimensionless)
$C_{\rm a}$	Atmospheric CO ₂ concentration (μmol mol ⁻¹)
$C_{\rm c}$	CO ₂ concentration at the site of Rubisco (µmol mol ⁻¹)
C_{i}	Intercellular CO ₂ concentration (µmol mol ⁻¹)
$C_{\rm r}$	LI-COR, (1990, 2006) reference CO ₂ concentration entering the
	measurement chamber (μmol mol ⁻¹)
$C_{\rm s}$	LI-COR, (1990, 2006) sample CO ₂ concentration exiting the
	measurement chamber (μmol mol ⁻¹)
$D_{\rm o}$	Empirical coefficient (2 kPa)
D_{s}	Vapor pressure deficit at the leaf surface (kPa)
$\Delta H_{\rm a}$	Activation energy (J mol ⁻¹)
$\Delta H_{ m d}$	Deactivation energy (J mol ⁻¹)
E	Transpiration rate per unit leaf area (mol m ⁻² s ⁻¹)
g_{m}	CO ₂ transfer conductance from the intercellular airspaces of the
	leaf into the chloroplast, referred to as mesophyll conductance
	$(\text{mol m}^{-2} \text{ s}^{-1})$
\overline{g}_{m}	Mean value of g _m (mol m ⁻² s ⁻¹)
g_{s}	Stomatal conductance to water vapor (mol m ⁻² s ⁻¹)
$g_{ m tc}$	Total conductance to CO ₂ (mol m ⁻² s ⁻¹)
Γ^*	CO ₂ compensation point in the absence of dark respiration
	(μmol mol ⁻¹)
J	Rate of electron transport (μmol m ⁻² s ⁻¹)
$J(g_{\rm m})$	Estimated J based on finite $g_m(\mu mol m^{-2} s^{-1})$.
J(∞)	Estimated J when g_m is assumed to be infinite (μ mol m ⁻² s ⁻¹).
$J_{ m max}$	Maximum rate of electron transport (μmol m ⁻² s ⁻¹)
$J_{\text{max}}(g_{\text{m}})$	Estimated J_{max} based on finite $g_{\text{m}}(\mu\text{mol m}^{-2}\text{ s}^{-1})$
$J_{\max}(\infty)$	Estimated J_{max} when g_{m} is assumed to be infinite (µmol m ⁻² s ⁻¹)
K_{c}	Michaelis constant for CO ₂ (mmol mol ⁻¹)
K_{o}	Michaelis constant for O ₂ (mmol mol ⁻¹)
$O_{\rm i}$	Intercellular oxygen concentration (210 mmol mol ⁻¹)
Q	Photosynthetically active photon flux density (μmol m ⁻² s ⁻¹)
Q_2	Irradiance absorbed by photosystem II (μmol m ⁻² s ⁻¹)
R	Molar gas constant (8 314 J K ⁻¹ mol ⁻¹)
$R_{\rm d}$	Dark (mitochondrial) respiration (μmol m ⁻² s ⁻¹)
T_{c}	Leaf temperature in °C
$T_{\mathbf{k}}$	Leaf temperature in Kelvins
$T_{ m opt}$	Optimal leaf temperature in Kelvins
$V_{\rm cmax}$	Maximum rate of Rubisco carboxylation (μmol m ⁻² s ⁻¹)
$V_{\rm cmax}(g_{\rm m})$	Estimated V_{cmax} based on finite g_{m} (μ mol m ⁻² s ⁻¹)
$V_{\rm cmax}(\infty)$	Estimated V_{cmax} when g_{m} is assumed to be infinite (µmol m ⁻² s ⁻¹)

References

Arkebauer, T.J., 1998. BOREAS TE-12 Leaf Gas Exchange Data. Data Set. Available on-line. Oak Ridge National Laboratory Distributed Active Archive Center, Oak Ridge, Tennessee, USA. http://www.daac.ornl.gov fromdoi:10.3334/ ORNLDAAC/351.

Ball, J.T., Woodrow, I.E., Berry, J.A., 1987. A model predicting stomatal conductance and its contribution to the control of photosynthesis under different environmental conditions. In: Biggens, J. (Ed.), Progress in Photosynthesis Research. Martinus Nijhoff Publishers, The Netherlands.

Bernacchi, C.J., Singsaas, E.L., Pimentel, C., Portis, A.R., Long, S.P., 2001. Improved temperature response functions for models of Rubisco-limited photosynthesis. Plant. Cell and Environment 24. 253–259.

Bernacchi, C.J., Portis, A.R., Nakano, H., von Caemmerer, S., Long, S.P., 2002. Temperature response of mesophyll conductance. Implications for the determination of Rubisco enzyme kinetics and for limitations to photosynthesis in vivo. Plant Physiology 130, 1992–1998.

Bernacchi, C.J., Pimentel, C., Long, S.P., 2003. In vivo temperature response functions of parameters required to model RuBP-limited photosynthesis. Plant, Cell and Environment 26, 1419–1430.

von Caemmerer, S., 2000. Techniques in Plant Sciences Volume 2: Biochemical Models of Leaf Photosynthesis. CSIRO Publishing, Collingwood.

Davey, P.A., Hunt, S., Hymus, G.J., DeLucia, E.H., Drake, B.G., Karnosky, D.F., Long, S.P., 2004. Respiratory oxygen uptake is not decreased by an instantaneous elevation of [CO₂], but is increased with long-term growth in the field at elevated [CO₂]. Plant Physiology 134, 520–527.

Dewar, R.C., 2002. The Ball-Berry-Leuning and Tardieu-Davies stomatal models: synthesis and extension within a spatially aggregated picture of guard cell function. Plant, Cell and Environment 25, 1383–1398.

- Dickson, R.E., Lewin, K.F., Isebrands, J.G., Coleman, M.D., Heilman, W.E., Riemenschneider, D.E., Sôber, J., Host, G.E., Zak, D.R., Hendrey, G.R., Pregitzer, K.S., Karnosky, D.F., 2000. Forest Atmosphere Carbon Transfer and Storage (FACTS-II) the Aspen Free-air CO₂ and O₃ Enrichment (FACE) Project: an Overview. Gen Tech. Rep. NC-214. Department of Agriculture, Forest Service, North Central Research Station, St. Paul, MN: U.S.
- Dubois, J.J.B., Fiscus, E.L., Booker, F.L., Flowers, M.D., Reid, C.D., 2007. Optimizing the statistical estimation of the parameters of the Farquhar-von Caemmerer-Berry model of photosynthesis. New Phytologist 176, 402–414.
- Ethier, G.J., Livingston, N.J., 2004. On the need to incorporate sensitivity to CO₂ transfer conductance into the Farquhar-von Caemmerer-Berry leaf photosynthesis model. Plant, Cell and Environment 27, 137–153.
- Farquhar, G.D., von Caemmerer, S., 1982. Modeling of photosynthetic responses to environmental conditions. In: Lange, O.L., Nobel, P.S., Osmond, C.B., Ziegler, H. (Eds.), Encyclopedia of Plant Physiology (New Series). Springer-Verlag, Berlin, pp. 549–587.
- Farquhar, G.D., von Caemmerer, S., Berry, J.A., 1980. A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species. Planta 149, 78–90.
- Farquhar, G.D., von Caemmerer, S., Berry, J.A., 2001. Models of photosynthesis. Plant Physiology 125, 42–45.
- Farquhar, G.D., Sharkey, T.D., 1982. Stomatal conductance and photosynthesis. Annual Reviews of Plant Physiology 33, 317–345.
- Flexas, J., Diaz-Espejo, A., Galmes, J., Kaldenhoff, R., Medrano, H., Ribas-Carbo, M., 2007. Rapid variations of mesophyll conductance in response to changes in CO₂ concentration around leaves. Plant, Cell and Environment 30, 1284–1298
- Flexas, J., Ribas-Carbo, M., Diaz-Espejo, A., Galmes, J., Medrano, H., 2008. Mesophyll conductance to CO₂: current knowledge and future prospects. Plant, Cell and Environment 31, 602–621.
- Harley, P.C., Loreto, F., Di Marco, G., Sharkey, T.D., 1992a. Theoretical considerations when estimating the mesophyll conductance to CO₂ flux by analysis of the response of photosynthesis to CO₂. Plant Physiology 98, 1429–1436.
- Harley, P.C., Thomas, R.B., Reynolds, J.F., Strain, B.R., 1992b. Modeling photosynthesis of cotton grown in elevated CO₂. Plant, Cell and Environment 15, 271–282.
- Harley, P.C., Tenhunen, J.D., 1991. Modeling the photosynthetic response of C_3 leaves to environmental factors. In: Boote, K.J., Loomis, R.S. (Eds.), Modeling Crop Photosynthesis from Biochemistry to Canopy. Crop Science of America, Madison, pp. 17–39.
- Hikosaka, K., Ishikawa, K., Borjigidal, A., Muller, O., Onoda, Y., 2006. Temperature acclimation of photosynthesis: mechanisms involved in the changes in temperature dependence of photosynthetic rate. Journal of Experimental Botany 57 (No. 2), 291–302.
- Jahnke, S., 2001. Atmospheric CO₂ concentration does not directly affect leaf respiration in bean or poplar. Plant, Cell and Environment 24, 1139–1151.
- Karnosky, D.F., Zak, D.R., Pregitzer, K.S., Awmack, C.S., Bockheim, J.G., Dickson, R.E., Hendrey, G.R., Host, G.E., Jones, W.S., King, J.S., Kopper, B.J., Kruger, E.L., Kubiske, M.E., Lindroth, R.L., Mattson, W.J., McDonald, E.P., Noormets, A., Oksanen, E., Parsons, W.F.J., Percy, K.E., Podila, G.K., Riemenschneider, D.E., Sharma, P., Sôber, A., Sôber, J., Anttonen, S., Vapaavuori, E., Isebrands, J.G., 2003. Tropospheric O₃ moderates responses of temperate hardwood forests to elevated CO₂: a synthesis of molecular to ecosystem results from the Aspen FACE project. Functional Ecology 17, 289–304.
- Kim, S.H., Lieth, J.H., 2003. A coupled model of photosynthesis, stomatal conductance and transpiration for a rose leaf (Rosa hybrida L.). Annals of Botany 91, 771–781.
- Kubiske, M.E., Zak, D.R., Pregitzer, K.S., Takeuchi, Y., 2002. Photosynthetic acclimation of overstory *Populus tremuloides* and understory *Acer saccharum* to elevated atmospheric CO₂ concentration: interactions with shade and soil nitrogen. Tree Physiology 22, 321–329.
- Kull, O., Sôber, A., Coleman, M.D., Dickson, R.E., Isebrands, J.G., Gagnon, Z., Karnosky, D.F., 1996. Photosynthetic responses of aspen clones to simultaneous exposures of O₃ and CO₂. Canadian Journal of Forestry Research 26, 639–648.
- Leuning, R., 1995. A critical appraisal of a combined stomatal-photosynthesis model for C_3 plants. Plant, Cell and Environment 18, 339–355.
- LI-COR, 1990. LI-6200 Technical Reference Manual. LI-COR, Inc, Lincoln, NE, USA. LI-COR, 1998. Using the LI-6400 Portable Photosynthesis System, second ed. LI-COR,
- Inc, Lincoln, NE, USA.
 LI-COR, 2006. Using the LI-6400 Portable Photosynthesis System. Version 5. http://www.licor.com/env/Products/li6400/6400_manuals.jsp

- Long, S.P., Bernacchi, C.J., 2003. Gas exchange measurements, what can they tell us about the underlying limitations to photosynthesis? Procedures and sources of error. Journal of Experimental Botany 54 (No. 392), 2393–2401.
- Long, S.P., Farage, P.K., Garcia, R.L., 1996. Measurement of leaf and canopy photosynthetic CO₂ exchange in the field. Journal of Experimental Botany 47, 1629–1642.
- Martin, M.J., Farage, P.K., Humphries, S.W., Long, S.P., 2000. Can the stomatal changes caused by acute ozone exposure be predicted by changes occurring in the mesophyll? A simplification for models of vegetation response to the global increase in tropospheric elevated ozone episodes. Australian Journal of Plant Physiology 27, 211–219.
- Martin, M.J., Host, G.E., Lenz, K.E., Isebrands, J.G., 2001. Simulating the growth response of aspen to elevated ozone: a mechanistic approach to scaling a leaf-level model of ozone effects on photosynthesis to a complex canopy architecture. Environmental Pollution 115. 425–436.
- McDermitt, D.K., Norman, J.M., Davis, J.T., Ball, T.M., Arkebaurer, T.J., Welles, J.M., Roemer, S.R., 1989. CO₂ response curves can be measured with a field-portable closed-loop photosynthesis system. Annals of Forestry Sciences (Suppl. 46), 416s–420s.
- Medlyn, B.E., Dreyer, E., Ellsworth, D., Forstreuter, M., Harley, P.C., Kirschbaum, M.U.F., Le Roux, X., Montpied, P., Strassemeyer, J., Walcroft, A., Wang, K., Loustau, D., 2002. Temperature response of parameters of a biochemically based model of photosynthesis. II. A review of experimental data. Plant, Cell and Environment 25, 1167–1179.
- Mott, K.A., Buckley, T.N., 2000. Patchy stomatal conductance: emergent collective behaviour of stomata. Trends in Plant Science 5 (6), 258–262.
- Niinemets, Ü, Oja, V., Kull, O., 1999. Shape of leaf photosynthetic electron transport versus temperature response curve is not constant along canopy light gradients in temperate deciduous trees. Plant, Cell and Environment 22. 1497–1513.
- Niinemets, Ü, Cescatti, A., Rodeghiero, M., Tosens, T., 2005. Leaf internal diffusion conductance limits photosynthesis more strongly in older leaves of Mediterranean evergreen broad-leaved species. Plant, Cell and Environment 28, 1552–1566.
- Noormets., A., Sôber, A., Pell, E.J., Dickson, R.E., Podila, G.K., Sôber, J., Isebrands, J.G., Karnosky, D.F., 2001a. Stomatal and non-stomatal limitation to photosynthesis in two trembling aspen (*Populus tremuloides Michx.*) clones exposed to elevated CO₂ and/or O₃. Plant, Cell and Environment 24, 327–336.
- Noormets, A., McDonald, E.P., Dickson, R.E., Kruger, E.L., Sôber, A., Isebrands, J.G., Karnosky, D.F., 2001b. The effect of elevated carbon dioxide and ozone on leafand branch-level photosynthesis and potential plant-level carbon gain in aspen. Trees 15, 262–270.
- Noormets, A., Kull, O., Sôber, A., Kubiske, M.E., Karnosky, D.F., 2010. Elevated CO₂ response of photosynthesis depends on ozone concentration in aspen. Environmental Pollution 158, 992–999.
- Pons, T.L., Welschen, R.A.M., 2002. Overestimation of respiration rates in commercially available clamp-on leaf chambers. Complications with measurement of net photosynthesis. Plant, Cell and Environment 25, 1367–1372.
- Reich, P.B., 1983. Effects of low concentration of O₃ on net photosynthesis, dark respiration, and chlorophyll contents in aging hybrid poplar leaves. Plant Physiology 73, 291–296.
- Rodeghiero, M., Niinemets., Ü, Cescatti, A., 2007. Major diffusion leaks of clamp-on leaf cuvettes still unaccounted: how erroneous are the estimates of Farquhar et al. model parameters? Plant, Cell and Environment 30, 1006–1022.
- Rogers, A., Humphries, S.W., 2000. A mechanistic evaluation of photosynthetic acclimation at elevated CO₂. Global Change Biology 6, 1005–1011.
- Schultz, H.R., 2003. Extension of a Farquhar model for limitations of leaf photosynthesis induced by light environment, phenology and leaf age in grapevines (*Vitis vinifera* L. cvv. White Riesling and Zinfandel). Functional Plant Biology 30, 673–687.
- Schurr, U., Walter, A., Rascher, U., 2006. Functional dynamics of plant growth and photosynthesis from steady-state to dynamics from homogeneity to heterogeneity. Plant, Cell and Environment 29, 340–352.
- Sharkey, T.D., 1985. Photosynthesis in intact leaves of C₃ plants: physics, physiology and rate limitations. The Botanical Review 51, 53–105.
- Sharkey, T.D., Bernacchi, C.J., Farquhar, G.D., Singsaas, E.L., 2007. Fitting photosynthetic carbon dioxide response curves for C₃ leaves. Plant, Cell and Environment 30, 1035–1040.
- Singsaas, E.L., Ort, D.R., Delucia, E.H., 2003. Elevated CO₂ effects on mesophyll conductance and its consequences for interpreting photosynthetic physiology. Plant, Cell and Environment 27, 41–50.